

## Antioxidant Effect of Two Virgin Olive Oils Depends on the Concentration and Composition of Minor Polar Compounds

FLAVIA FRANCONI,<sup>†,‡</sup> RITA COINU,<sup>\*,#</sup> STEFANIA CARTA,<sup>†</sup>  
 PIETRO PAOLO URGEGHE,<sup>†</sup> FRANCESCA IERI,<sup>§</sup> NADIA MULINACCI,<sup>§</sup> AND  
 ANNALISA ROMANI<sup>§</sup>

Centre for Biotechnology Development and Biodiversity Research, University of Sassari, via Muroli 25, 07100 Sassari, Italy; Department of Physiological, Biochemical, and Cellular Science, University of Sassari, Sassari, Italy; Department of Pharmacology, University of Sassari, Sassari Italy; and Department of Pharmaceutical Science, University of Florence, Sesto F.no (FI), Italy

*In vitro* studies show that some individual minor polar phenolic compounds (MPC) present in virgin olive oil prevent oxidation of human low-density lipoproteins (LDL), but few data are available on the antioxidant effect of whole oil extract. Thus, whole virgin olive extracts were studied to determine whether they maintain the antioxidant activity and whether this last is linked to MPC composition of a single virgin oil. Using HPLC-DAD the MPC content in Taggiasca and Seggianese virgin olive oils was measured. Taggiasca oil was less rich in total MPC (208.5 mg/L) than Seggianese oil (441.9 mg/L). In addition, the major compounds of Taggiasca oil were lignan derivatives, whereas the major compounds in Seggianese oils were secoiridoid derivatives. Moreover, Taggiasca oil was practically free of 5-hydroxytyrosol and 5-hydroxytyrosol derivatives, deacetoxy-oleuropein aglycone and oleuropein aglycone. The antioxidant activity of the oils on human LDL was evaluated by measuring malondialdehyde and conjugate diene generation induced by copper ions. In both tests, the oil extracts dose-dependently reduced malondialdehyde and conjugate diene generation. Moreover, antioxidant potency correlated with total MPC; thus, Seggianese extract was more active. The two oils differed quantitatively and qualitatively, and these differences influenced their biological activities; thus clinical trials focused on studying the effects of olive oils should specify the oils used.

**KEYWORDS:** Antioxidant activity; LDL; MDA; secoiridoids; lignans; deacetoxy-oleuropein; acetoxy-pinoresinol; virgin olive oil

### INTRODUCTION

Epidemiological studies suggest that Mediterranean diets are associated with a reduced risk of cardiovascular diseases, the lower incidence of cardiovascular disease being associated with greater adherence to the Mediterranean diet (1–3). Recently, it has been shown in more than 3000 individuals, without clinical evidence of cardiovascular disease, that total plasma antioxidant capacity and low oxidized low-density lipoproteins (LDL) are associated with greater adherence to the Mediterranean diet (4). Compared with a saturated fat diet, the Mediterranean diet, rich in oleic acid, has also been associated with lower LDL and total triglycerides with maintenance of high-density lipoproteins (5–7) and lower blood pressure (8, 9).

Olive oil composition depends on many factors such as olive cultivar (10–12) and agronomic and technological aspects of production (13–15). Virgin oil, which is obtained directly from pressing ripe olives, retains sizable amounts of minor polar compounds (MPC) and tocopherols (16), which can act as antioxidants (17). Recently, it was shown that the antioxidant activity of monovarietal extra virgin olive oils was increased in those oils extracted from destoned fruits and that this effect was variety-dependent (18).

It is known that some single MPC increase the resistance of LDL against oxidation *in vitro* (19–23), but the single MPC approach fails to account for the interactions among MPC and does not take into consideration that some MPC are correlated between them. In this respect, it is important to note that the mixture of phenols may exert different activity in comparison with the single phenols, because they may cooperate, thereby modifying biological activity (24). Interactions among phenols seems also to depend on the relative amount of single polyphenols (25). Thus, individual olive oils, which differ qualitatively and quantitatively, could have different biological activities. At present, little is known about the antioxidant effect of the total

\* Corresponding author (telephone +39 79228654; fax +39 79228615; e-mail coinu@uniss.it).

<sup>†</sup> Centre for Biotechnology Development and Biodiversity Research, University of Sassari.

<sup>#</sup> Department of Physiological, Biochemical, and Cellular Science, University of Sassari.

<sup>‡</sup> Department of Pharmacology, University of Sassari.

<sup>§</sup> University of Florence.

olive oil extract. In addition, the relationship between antioxidant activity of whole virgin olive oil and its global MPC content is not yet known (26, 27). The foregoing observations may account for the conflicting results obtained in clinical trials designed to determine the effect of olive oil (27–31). Interestingly, in two of these studies, the beneficial effects of virgin olive oil were related to MPC amount (27, 30). Olive oil is a basic component of the Mediterranean diet; thus, it is important to identify and quantify the antioxidant compounds in individual virgin oils so as to determine which have the healthiest effects.

## MATERIALS AND METHODS

Tyrosol, luteolin, and oleuropein were obtained from Extrasynthèse (Genay, France). 5-Hydroxytyrosol was purchased from Cayman Chemical (SPI-BIO, Montigny le Bretonneux, France). Solvents for the high-performance liquid chromatography–diode array detection (HPLC-DAD) analyses were of analytical grade and were purchased from Carlo Erba (Milan, Italy). Other reagents were of analytical grade and were purchased from Sigma (St. Louis, MO).

**Preparation, Characterization, and Quantification of Virgin Olive Oil Extracts.** The virgin oils were produced from two autochthonous cultivars: ‘Olivastra Seggianese’ (Tuscany, Italy) and ‘Taggiasca’ (Liguria, Italy). The olives were processed immediately after harvesting by Manni S.p.A. (Grosseto, Italy) and Isnardi S.p.A. (Imperia, Italy), respectively. Sample preparation and extraction and identification, characterization, and quantification of single polar compounds were carried out as previously reported (32). Briefly, 50 mL of each fresh oil sample was extracted with 150 mL of ethanol and formic acid acidified water (70:30 v/v). Defatting with *n*-hexane was carried out to remove the lipid fraction. The extract was concentrated under reduced pressure to dryness, dissolved with 2 mL of extraction solvent, and analyzed by HPLC using an HP-1100 liquid chromatograph equipped with a DAD detector and an HP 1100 MSD API–electrospray (Agilent Technologies, Palo Alto, CA). The MPC were identified on the basis of their retention times and spectroscopic and spectrometric data, using 5-hydroxytyrosol, tyrosol, luteolin, and oleuropein as reference compounds. Lignan was identified and analyzed as described in Mulinacci et al. (33). Oleocanthal was identified according to the method of Beauchamp et al. (34). The single minor compounds were quantified with HPLC-DAD using a four-point regression curve constructed with the available standards. Calibration curves with  $r^2 \geq 0.9998$  were used (11). In all cases actual concentrations of derivatives were calculated after the application of corrections for changes in molecular weight: knowing the molecular weight of each compound ( $PM_x$ ) allowed its actual concentration to be obtained by applying a multiplication factor of  $PM_x/PM_y$ , where  $PM_y$  is the molecular weight of the specific reference compound. The same extract analyzed by HPLC was used for the LDL test.

**Subjects.** Thirty healthy, sex-matched volunteers gave their informed consent to participate in the study. Smokers were excluded from the study. No subject had a family history of diabetes, hypertension, or dyslipidemia. They remained free of drugs, vitamins, amino acids, hormones, dietary supplements, and botanical remedies for the 14 days preceding the study. All of the subjects regularly consume a typical Mediterranean diet.

**LDL Isolation and Preparation.** In the morning after a 12-h fast, 40 mL of blood was collected by venipuncture into EDTA-containing vacutainer tubes (1 g/L) and centrifuged immediately for 10 min at 2000g at 4 °C. LDL was isolated from plasma with the discontinuous ultracentrifugation method using a TL-100 tabletop ultracentrifuge (Beckman, Palo Alto, CA) as reported previously (35) with minor revisions. To protect the LDL against oxidative alterations during ultracentrifugation, each density solution contained EDTA. The samples were then exhaustively dialyzed in PD-10 desalting columns (Amersham Pharmacia Biotech, Uppsala, Sweden) to remove excess salt and most of the EDTA. The LDL samples were used immediately thereafter.

**Biochemical Assays.** Total cholesterol, LDL, high-density lipoprotein (HDL), and triglycerides were measured as previously described (36). The protein concentration of LDL was determined using bovine

serum albumin as standard (37). LDL was diluted to 50  $\mu$ g of protein/mL, and LDL oxidation was carried out as previously described in Romani et al. (25). Oxidation was initiated by adding freshly prepared 5  $\mu$ M  $\text{CuSO}_4$  (final concentration). LDL oxidation was measured in triplicate with the thiobarbituric acid method, using malonaldehyde bisdiethylacetal as standard (38). The malondialdehyde was measured in basal condition and after 8 h of exposure to copper ions, in the presence and absence of different concentrations of extra virgin oil extracts added before copper ions. Conjugated diene formation was determined at a temperature of 37 °C and by monitoring the absorbance at 234 nm, as described elsewhere (39).

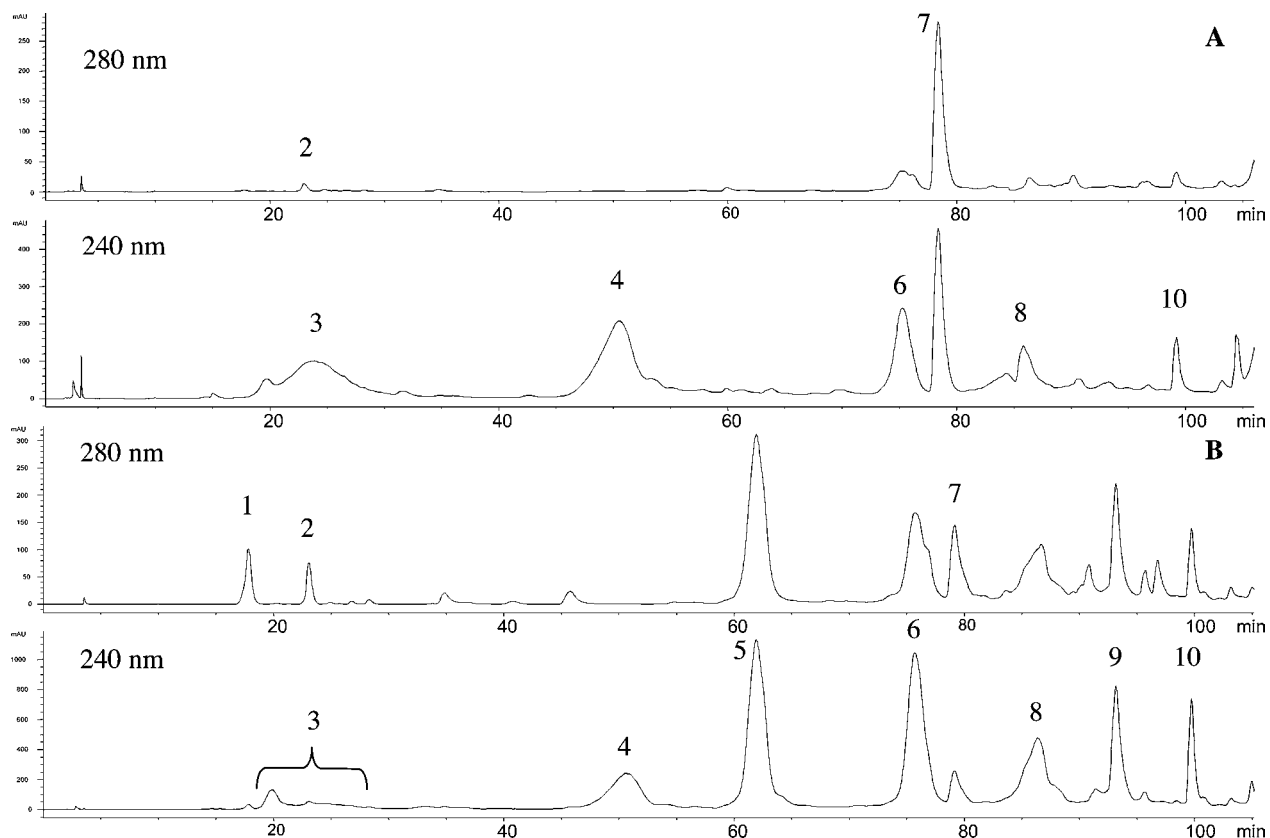
**Statistical Analysis.** The results are expressed as mean  $\pm$  standard error (SE).  $\text{IC}_{50}$  was calculated using the Sigmaplot 8.0 program (SPSS Inc., Chicago, IL). ANOVA followed by Turkey’s multiple-range test was used to compare means at a significance level of  $P < 0.05$ .

## RESULTS AND DISCUSSION

As shown in **Figure 1** and **Table 1**, the MPC identified and quantified in the two virgin olive oils belong to three classes: simple phenols (tyrosol, 5-hydroxytyrosol, and 5-hydroxytyrosol derivatives), secoiridoids (oleuropein aglycones, deacetoxy-oleuropein aglycone, oleocanthal, and secoiridoids derivatives), and lignan derivatives (acetoxypinoresinol). The Taggiasca extract contained a high percentage of lignan derivatives (72.5%) and no 5-hydroxytyrosol, 5-hydroxytyrosol derivatives, deacetoxy-oleuropein aglycone, or oleuropein aglycone (**Figure 1** and **Table 1**). The Seggianese extract contained tyrosol, 5-hydroxytyrosol, 5-hydroxytyrosol derivatives, deacetoxy-oleuropein aglycone, and oleuropein aglycone. In addition, it was rich in secoiridoids and had a relatively low amount of lignan derivatives. Both oil extracts contain oleocanthal, the Saggianese cultivar being richer than the Taggiasca (**Figure 1** and **Table 1**). 5-Hydroxytyrosol and oleuropein aglycone show strong antioxidant properties (40). Oleocanthal is reported to be an inhibitor of cyclooxygenases 1 and 2 (34); on the other hand, there are no studies about antioxidant properties of deacetoxy-oleuropein aglycone as a single compound. Acetoxypinoresinol shows only low antioxidant properties (41), and to our knowledge tyrosol and elenolic acid do not possess any antioxidant properties (42).

None of the extracts affected the basal level of malondialdehyde (data not shown) in LDL from healthy individuals. Hence, we evaluated the effect of the oil extracts on malondialdehyde production induced by copper ions in human LDL. We found that the olive oils dose-dependently inhibited malondialdehyde generation, and the  $\text{IC}_{50}$  showed that the Seggianese extract was the most potent when total MPC were considered. The shapes of dose–response curves differed, but when efficiency was calculated, the difference was not significant (data not shown). Measurable LDL conjugated diene formation did not occur in the absence of copper ions (data not shown). Oil extracts shifted the conjugated diene formation curve to the right (**Figure 2**).

Diet is a cornerstone of cardiovascular disease prevention (43), and epidemiological studies demonstrate that Mediterranean populations have a lower incidence of cardiovascular disease (44). The healthy effect of the Mediterranean diet has been attributed to the consumption of large amounts of fiber, fruits, vegetables, and unsaturated fatty acid provided by olive oil. However, the contribution of virgin olive oil as a major source of dietary antioxidants has not been investigated in depth (45). Our data confirm that virgin olive oils contain antioxidants in abundance (11, 32). The level of MPC was about 2.1 higher in Saggianese oil extract than in the Taggiasca one, which indicates that the cultivar used to produce the oil is an important



**Figure 1.** HPLC-DAD profiles of (A) Taggiasca virgin oil extract and (B) Seggianese virgin oil extract acquired at 280 and 240 nm. Identified compounds: 1, 5-hydroxytyrosol; 2, tyrosol; 3, elenolic acid derivatives; 4, elenolic acid; 5, deacetoxy-oleuropein aglycone; 6, oleocanthal; 7, acetoxypinoresinol; 8, oleuropein aglycone; 9–10, secoiridoids.

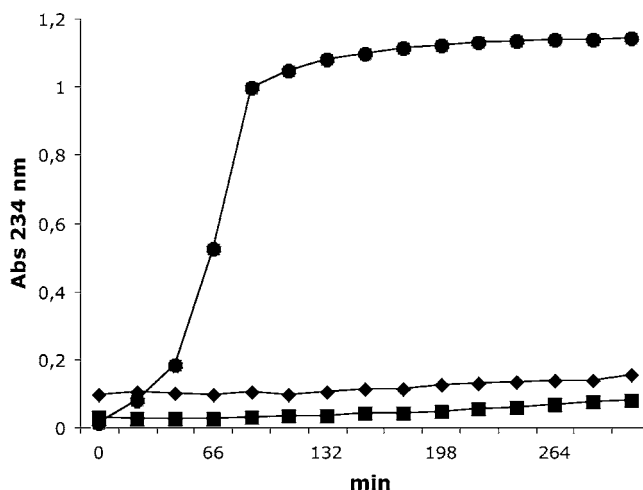
**Table 1.** Composition of Virgin Oil Extracts<sup>a</sup>

compound	Taggiasca oil, mg/L (%)	Seggianese oil, mg/L (%)
tyrosol	1.5 (0.7)	9.5 (2.1)
5-hydroxytyrosol	traces	14.0 (3.2)
5-hydroxytyrosol derivatives	ND	2.4 (0.5)
oleocanthal	8.3 (4.0)	53.0 (12.0)
elenolic acid	26.8 (12.9)	28.6 (6.5)
elenolic acid derivatives	15.8 (7.6)	6.6 (1.5)
deacetoxy-oleuropein aglycone	ND	102.1 (23.1)
oleuropein aglycones	ND	50.8 (11.5)
secoiridoid derivatives	3.6 (1.7)	51.0 (11.5)
lignan derivatives <sup>b</sup>	151.2 (72.5)	122.0 (27.6)
luteolin	1.3 (0.6)	1.9 (0.4)
total polyphenols	208.5 (100)	441.9 (100)

<sup>a</sup> Data reported are the mean of three determinations each performed in triplicate; SE was in the range 1–3%; ND, nondetectable. <sup>b</sup> Mainly acetoxypinoresinol.

factor in determining the MPC content. This observation is in line with earlier studies indicating that the cultivar, as well as agronomic and technological aspects of production, is related to MPC levels (11, 14, 32).

The two olive oil extracts examined in our study differ not only in total MPC levels but also in chemical composition. In the Taggiasca extract, the major compounds were lignans (72.5%) that are mainly constituted by pinoresinol derivatives (45), which seem to have a low antioxidant activity (41), whereas, in the Seggianese extract, the major compounds were secoiridoids (deacetoxy-oleuropein aglycone, 23.1%; oleuropein aglycone, 11.5%; and secoiridoid derivatives, 11.5%), which have high antioxidant activity. In addition, both oils contained different amounts of oleocanthal, an inhibitor of the prosta-



**Figure 2.** Typical profiles of conjugate diene formations induced by copper ions in human LDL (●); inhibitory effect of Taggiasca (◆) and Seggianese (■) oil extracts. Both Taggiasca and Seggianese extracts were used at a concentration of 5  $\mu$ M as MPC.

glandin-biosynthesis pathway (34) suggestive of anti-inflammatory activity, whereas flavonol levels did not differ substantially between the two oils. Interestingly, the two oil extracts also contain MPC that do not exert antioxidant activity (Figure 3), that is, tyrosol, elenolic acid, and derivatives of elenolic acid, and their total amounts are more elevated in Seggianese than in Taggiasca. Finally, the Taggiasca oil extract lacks 5-hydroxytyrosol, 5-hydroxytyrosol derivatives, deacetoxy-oleuropein aglycone, and oleuropein aglycone, whereas all of these compounds are present in the Seggianese oil extract.

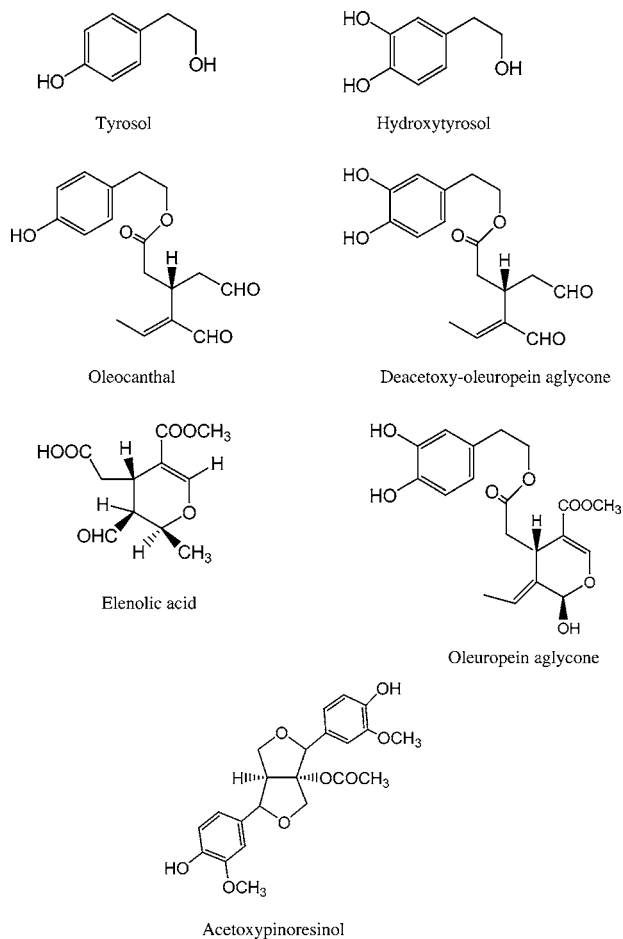


Figure 3. Structures of MPC.

Theoretically, the extract of Seggianese virgin oil, which has a greater amount of MPC and a lesser amount of MPC lacking in antioxidant activity, would exert a strong antioxidant effect. Indeed, the Seggianese oil extract was more potent ( $\sim 4$  times) compared with the Taggiasca oil extract. Importantly, the antioxidant effects and therefore the possible antiatherogenic activities of the two virgin oils were dose-dependent and occur at low concentrations:  $IC_{50}$  values of  $2.1 \pm 0.9$  and  $0.6 \pm 0.2$   $\mu M$  for Taggiasca and Seggianese oil extracts, respectively. After nutritional intake of virgin olive oil, the plasma level of MPC is  $\sim 0.6$   $\mu M$  (46), which is similar to the  $IC_{50}$  calculated for Seggianese oil extracts.

In conclusion, there are notable quantitative and qualitative differences between the two oil extracts studied, which could explain the differences observed in their antioxidant activities. In a small clinical study carried out with olive oils containing different amounts of MPC, the short-term inhibition of LDL oxidation is greater with oils rich in MPC (30). On the basis of this finding and our data, we suggest that clinical trials designed to evaluate the healthy properties of virgin olive oils should specify the type of virgin oil being tested and possibly its chemical composition, or at least the amount of MPC contained in the oil, so that results from different studies can be compared. It seems that the presence of 5-hydroxytyrosol and its derivatives and high levels of secoiridoids enhance antioxidant activities, suggesting that virgin olive oil rich in these compounds could have more potent health-protecting properties because the antioxidant activity of oils could have beneficial effect according to the oxidative hypothesis of atherosclerosis (47–50).

## ABBREVIATIONS USED

MPC, minor polar phenolic compounds; LDL, human low-density lipoproteins; HPLC-DAD, high-performance liquid chromatography–diode array detection.

## LITERATURE CITED

- Keys, A.; Menotti, A.; Karvonen, M. J.; Aravanis, C.; Blackburn, H.; Buzina, R.; Djordjevic, B. S.; Dontas, A. S.; Fidanza, F.; Keys, M. H. The diet and 15-year death rate in the seven countries study. *Am. J. Epidemiol.* **1986**, *124*, 903–915.
- Cuevas, A. M.; Germain, A. M. Diet and endothelial function. *Biol. Res.* **2004**, *372*, 225–230.
- Kok, F. J.; Kromhout, D. Atherosclerosis-epidemiological studies on the health effects of a Mediterranean diet. *Eur. J. Nutr.* **2004**, *43* (Suppl. 1), 2–5.
- Pitsavos, C.; Panagiotakos, D. B.; Tzima, N.; Chrysohoou, C.; Economou, M.; Zampelas, A.; Stefanadis, C. Adherence to the Mediterranean diet is associated with total antioxidant capacity in healthy adults: the ATTICA study. *Am. J. Clin. Nutr.* **2005**, *82*, 694–699.
- Grundy, S. M. Comparison of monounsaturated fatty acids and carbohydrates for lowering plasma cholesterol. *N. Engl. J. Med.* **1986**, *314*, 745–748.
- Mensink, R. P.; Katan, M. B. Effect of monounsaturated fatty acids versus complex carbohydrates on high-density lipoproteins in healthy men and women. *Lancet* **1987**, *8525*, 122–125.
- Aguilera, C. M.; Ramirez-Tortosa, M. C.; Mesa, M. D.; Ramirez-Tortosa, C. L.; Gil, A. Sunflower, virgin-olive and fish oils differentially affect the progression of aortic lesions in rabbits with experimental atherosclerosis. *Atherosclerosis* **2002**, *162*, 335–344.
- Ferro-Luzzi, A.; Strazzullo, P.; Scaccini, C.; Siani, A.; Sette, S.; Mariani, M. A.; Mastranzo, P.; Dougherty, R. M.; Iacono, J. M.; Mancini, M.; Changing the Mediterranean diet: effects on blood lipids. *Am. J. Clin. Nutr.* **1984**, *40*, 1027–1037.
- Strazzullo, P.; Ferro-Luzzi, A.; Siani, A.; Scaccini, C.; Sette, S.; Catata, G.; Mancini, M. Changing the Mediterranean diet: effects on blood pressure. *J. Hypertens.* **1986**, *4*, 407–412.
- Flamini, G.; Cioni, P. L.; Morelli, I. Volatiles from leaves, fruits, and virgin oil from *Olea europaea* cv. Olivastra Seggianese from Italy. *J. Agric. Food Chem.* **2003**, *51*, 1382–1386.
- Pinelli, P.; Galardi, C.; Mulinacci, N.; Romani, A. Minor polar compound and fatty acid analyses in monocultivar virgin olive oils from Tuscany. *Food Chem.* **2003**, *80*, 331–336.
- Kalua, C. M.; Allen, M. S.; Bedgood, D. R., Jr.; Bishop, A. G.; Prenzl, P. D. Discrimination of olive oils and fruits into cultivars and maturity stages based on phenolic and volatile compounds. *J. Agric. Food Chem.* **2005**, *53*, 8054–8062.
- Amirante, P.; Catalano, P.; Amirante, R.; Clodoveo, M. L.; Montel, G. L.; Leone, A.; Tamborrino, A. Prove sperimentali di estrazione di oli da olive snocciate. *Olive Olio* **2002**, *6*, 16–22.
- Servili, M.; Selvaggini, R.; Esposto, S.; Taticchi, A.; Montedoro, G.; Morozzi, G. Health and sensory properties of virgin olive oil hydrophilic phenols: agronomic and technological aspects of production that affect their occurrence in the oil. *J. Chromatogr. A* **2004**, *1054*, 113–127.
- Mulinacci, N.; Giaccherini, C.; Innocenti, M.; Romani, A.; Vincieri, F. F.; Marotta, F.; Mattei, A. Analysis of extra virgin olive oils from stoned olives. *J. Sci. Food Agric.* **2005**, *85*, 662–670.
- Visioli, F.; Galli, C. Antiatherogenic components of olive oil. *Curr. Atheroscler. Rep.* **2001**, *3*, 64–67.
- Mateos, R.; Dominguez, M. M.; Espartero, J. L.; Cert, A. Antioxidant effect of phenolics compounds,  $\alpha$ -tocopherol, and other minor components in virgin olive oil. *J. Agric. Food Chem.* **2003**, *51*, 7170–7175.

- (18) Lavelli, V.; Bondesan, L. Secoiridoids, tocopherols, and antioxidant activity of monovarietal extra virgin olive oils extracted from destoned fruits. *J. Agric. Food Chem.* **2005**, *53*, 1102–1107.
- (19) Manna, C.; Galletti, P.; Cucciolla, V.; Montedoro, G.; Zappia, V. Olive oil hydroxytyrosol protects human erythrocytes against oxidative damages. *J. Nutr. Biochem.* **1999**, *10*, 159–165.
- (20) O'Dowd, Y.; Driss, F.; Dang, P. M.; Elbim, C.; Gougerot-Pocidalo, M. A.; Pasquier, C.; El-Benna, J. Antioxidant effect of hydroxytyrosol, a polyphenol from olive oil: scavenging of hydrogen peroxide but not superoxide anion produced by human neutrophils. *Biochem. Pharmacol.* **2004**, *68*, 2003–2008.
- (21) Manna, C.; Migliardi, V.; Golino, P. Oleuropein prevents oxidative myocardial injury induced by ischemia and reperfusion. *J. Nutr. Biochem.* **2004**, *15*, 461–466.
- (22) Masella, R.; Vari, R.; D'Archivio, M.; Di Benedetto, R.; Matarrese, P.; Malorni, W.; Scazzocchio, B.; Giovannini, C. Extra virgin olive oil biophenols inhibit cell-mediated oxidation of LDL by increasing the mRNA transcription of glutathione-related enzymes. *J. Nutr.* **2004**, *134*, 785–791.
- (23) Moreno, J. J. Effect of olive oil minor components on oxidative stress and arachidonic acid mobilization and metabolism by macrophages RAW 264.7. *Free Radical Biol. Med.* **2003**, *35*, 1073–1081.
- (24) Reaven, P. D.; Witztum, J. L. Oxidized low-density lipoproteins in atherogenesis: role of dietary modification. *Annu. Rev. Nutr.* **1996**, *16*, 51–71.
- (25) Romani, A.; Coinu, R.; Carta, S.; Pinelli, P.; Galardi, C.; Vincieri, F. F.; Franconi, F. Evaluation of antioxidant effect of different extracts of *Myrtus communis* L. *Free Radical Res.* **2004**, *38*, 97–103.
- (26) Owen, R. W.; Haubner, R.; Wurtele, G.; Hull, E.; Spiegelhalter, B.; Bartsch, H. Olives and olive oil in cancer prevention. *Eur. J. Cancer Prev.* **2004**, *4*, 319–326.
- (27) Visioli, F.; Caruso, D.; Grande, S. Virgin Olive Oil Study (VOLOS): vasoprotective potential of extra virgin olive oil in mildly dyslipidemic patients. *Eur. J. Nutr.* **2004**, *6*, 1–7.
- (28) Nagyova, A.; Haban, P.; Klvanova, J.; Kadrabova, J. Effects of dietary extra virgin olive oil on serum lipid resistance to oxidation and fatty acid composition in elderly lipidemic patients. *Bratisl. Lek. Listy* **2003**, *104*, 218–221.
- (29) Marrugat, J.; Covas, M. I.; Fito, M.; Schroder, H.; Miro-Casas, E.; Gimeno, E.; Lopez-Sabater, M. C.; de la Torre, R.; Farre, M.; SOLOS Investigators. Effects of differing phenolic content in dietary olive oils on lipids and LDL oxidation—a randomized controlled trial. *Eur. J. Nutr.* **2004**, *43*, 140–147.
- (30) Weinbrenner, T.; Fito, M.; de la Torre, R.; Saez, G. T.; Rijken, P.; Tormos, C.; Coolen, S.; Albaladejo, M. F.; Abanades, S.; Schroder, H.; Marrugat, J.; Covas, M. I. Olive oils high in phenolic compounds modulate oxidative/antioxidative status in men. *J. Nutr.* **2004**, *134*, 2314–2321.
- (31) Rodríguez-Villar C.; Pérez-Heras A.; Mercadé I.; Casals E.; Ros. Comparison of a high-carbohydrate and a high-monounsaturated fat E., olive oil-rich diet on the susceptibility of LDL to oxidative modification in subjects with Type 2 diabetes mellitus. *Diabetes Med.* **2004**, *21*, 142–149.
- (32) Romani, A.; Pinelli, P.; Mulinacci, N.; Galardi, C.; Vincieri, F. F.; Liberatore, L.; Cichelli, A. HPLC and HRGC analyses of polyphenols and secoiridoids in olive oil. *Chromatographia* **2001**, *53*, 279–284.
- (33) Mulinacci, N.; Giaccherini, C.; Ieri, F.; Romani, A.; Vincieri, F. F. Evaluation of lignans and free and linked hydroxy-tyrosol and tyrosol in extra virgin olive oil after hydrolysis processes. *J. Sci. Food Agric.* **2006**, *86*, 757–764.
- (34) Beauchamp, G. K.; Keast, R. S. J.; Morel, D.; Lin, J.; Pika, J.; Han, Q.; Lee, C.-H.; Smith, A. B.; Breslin, P. A. S. Phytochemistry: ibuprofen-like activity in extra-virgin olive oil. *Nature* **2005**, *43*, 45–46.
- (35) Himber, J.; Bulher, E.; Moll, D.; Moser, U.K. Low-density lipoproteins for oxidation and metabolic studies. Isolation from small volumes of plasma using a tabletop ultracentrifuge. *Int. J. Vit. Nutr. Res.* **1995**, *65*, 137–142.
- (36) Franconi, F.; Miceli, M.; Alberti, L.; Boatto, G.; Coinu, R.; De Montis, M. G.; Tagliamonte, A. Effect of  $\gamma$ -hydroxybutyric acid on human platelet aggregation in vitro. *Thromb. Res.* **2001**, *102*, 255–260.
- (37) Bradford, M. M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein–dye binding. *Anal. Biochem.* **1976**, *72*, 248–254.
- (38) Esterbauer, H.; Cheeseman, K. H. Determination of aldehydic lipid peroxidation products: malonaldehyde and 4-hydroxynonenal. *Methods Enzymol.* **1990**, *186*, 407–421.
- (39) Esterbauer, H.; Striegl, G.; Rothender, M. Continuous monitoring of in vitro oxidation of human low-density lipoprotein. *Free Radical Res. Commun.* **1989**, *6*, 67–75.
- (40) Visioli, F.; Bellomo, G.; Galli, C. Free radical-scavenging properties of olive oil polyphenols. *Biochem. Biophys. Res. Commun.* **1998**, *247*, 60–64.
- (41) Nenadis, N.; Wang, L. F.; Tsimidou, M. Z.; Zhang, H. Y. Radical scavenging potential of phenolic compounds encountered in *O. europaea* products as indicated by calculation of bond dissociation enthalpy and ionization potential values. *J. Agric. Food Chem.* **2005**, *53*, 295–299.
- (42) Carluccio, M. A.; Siculella, L.; Ancora, M. A.; Massaro, M.; Scoditti, E.; Storelli, C.; Visioli, F.; Distanti, A.; De Caterina, R. Olive oil and red wine antioxidant polyphenols inhibit endothelial activation: antiatherogenic properties of Mediterranean diet phytochemicals. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 622–629.
- (43) Kris-Etherton, P.; Eckel, R. H.; Howard, B. V.; St Jeor, S.; Bazzarre, T. L. Lyon Diet Heart Study. Benefits of a Mediterranean-style diet. *Circulation* **2001**, *103*, 1823–1825.
- (44) Owen, R. W.; Haubner, R.; Wurtele, G.; Hull, E.; Spiegelhalter, B.; Bartsch, H. Olives and olive oil in cancer prevention. *Eur. J. Cancer Prev.* **2004**, *13*, 319–326.
- (45) Owen, R. W.; Mier, W.; Giacosa, A.; Hull, W. E.; Spiegelhalter, B.; Bartsch, H. Identification of lignans as major components in the phenolic fraction of olive oil. *Clin. Chem.* **2000**, *46*, 976–988.
- (46) Visioli, F.; Galli, Bornet, F.; Mattei, A.; Patelli, R.; Galli, G.; Caruso, D. Olive oil phenolics are dose-dependently absorbed in humans. *FEBS Lett.* **2000**, *468*, 159–160.
- (47) Steinberg, D.; Parthasarathy, S.; Carew, T. E.; Khoo, J. C.; Witztum, J. L. Beyond cholesterol. Modifications of low-density lipoprotein that increase its atherogenicity. *N. Engl. J. Med.* **1989**, *320*, 915–924.
- (48) Lusis, A. J. Atherosclerosis. *Nature* **2000**, *407*, 233–241.
- (49) Steinbrecher, U. P.; Loughheed, M. Scavenger receptor-independent stimulation of cholesterol esterification in macrophages by low-density lipoprotein extracted from human aortic intima. *Arterioscler. Thromb.* **1992**, *12*, 608–625.
- (50) Yla-Herttuala, S.; Palinski, W.; Rosenfeld, M. E.; Parthasarathy, S.; Carew, T. E.; Butler, S.; Witztum, J. L.; Steinberg, D. Evidence for the presence of oxidatively modified low-density lipoprotein in atherosclerotic lesions of rabbit and man. *J. Clin. Invest.* **1989**, *84*, 1086–1095.

Received for review December 1, 2005. Revised manuscript received February 22, 2006. Accepted February 22, 2006. This work was partly supported by MIUR Prin-2005055502\_003, Monte dei Paschi di Siena Foundation, and S.C. has a fellowship from GIO.I.A onlus.